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The Solid Phase Synthesis of Trisubstituted 1,4-Diazabicyclo[4.3.0]nonan-2-one Scaffolds: On Bead Monitoring of Heterocycle Forming Reactions Using ¹⁵N NMR

Eric E. Swayze

Isis Pharmaceuticals, 2292 Faraday Av., Carlsbad, CA 92008

Abstract: Several representative 3,4,8-trisubstituted 1,4-diazabicyclo[3.4.0]nonan-2-ones have been prepared employing solid phase methodologies. Elaboration of a 4-hydroxyproline derivative with an ¹⁵N-amino acid derivative allowed convenient monitoring of the reaction sequence on solid support by gel-phase ¹⁵N NMR. An intramolecular Mitsunobu cyclization provided the desired heterocycle, which could be further functionalized at the 4-position. This synthetic method is facile, general, and suitable for the construction of large libraries of compounds for biological assays. © 1997 Published by Elsevier Science Ltd.

In order to increase the diversity of new structures readily available for biological assays, we have undertaken studies to prepare a series of non-structure based molecules¹ amenable to assembly via combinatorial methods.² A particularly appealing strategy for accomplishing this goal involves the construction of a heterocyclic system using amino acids as key building blocks, since they are readily available from commercial suppliers, and bear functionality having a high degree of diversity. This approach was used by both the Ellman³ and Parke-Davis⁴ groups in their early work on the synthesis of small molecule combinatorial libraries in a exploration of known pharmacophores, and has more recently been applied to the synthesis of cyclic ureas,⁵ as well as novel heterocyclic structures.⁶ We envisioned a similar strategy, using amino acid derivatives as building blocks for the construction of the 1,4diazabicyclo[3.4.0]nonan-2-one ring system employing solid phase methodologies. Although the piperazinedione derivatives of this ring system have been more extensively investigated, derivatives unsubstituted at the 5-postion are unknown. A recent report in the literature⁷ describes the synthesis of a related fused piperazinone ring system by the intramolecular displacement of a mesyl group with an amine, however the yields were modest, and required forcing conditions. Since we are interested in the automated synthesis of a large number of compounds via solid phase methodologies for biological screening, we required facile reactions throughout the synthetic pathway in order to obtain acceptable yields and purities of the final products. These considerations led us to consider an intramolecular Mitsunobu reaction using the 2-nitrobenzenesulfonamide group⁸ as an easily removable activating group for effecting synthesis of the ring system.

As a model system, the protected *trans*-4-hydroxyproline derivative 1^9 (Scheme 1) was silylated, deprotected at nitrogen, then coupled to FMOC-Ala-OH to provide 2. The protecting group on the amino acid was then exchanged for the 2-nitobenzenesulfonyl group, and the DMT protection removed. Treatment of the resulting derivative 3 under Mitsunobu conditions effected a clean cyclization to the desired compound 4, providing a 99% yield of isolated product. The sulfonamide was then removed, the unsubstituted nitrogen functionalized by alkylation with 3-nitrobenzyl bromide, and the material desilylated to give 5.

The methodology was next adapted to the solid phase in order to facilitate synthesis of related structures on our automated high throughput parallel array synthesizer.¹⁰ A suitable attachment point to the support would be the free



Scheme 1. a) TBDPSCI/imidazole/DMF; b) 10% piperidine/DMF; c) FMOC-L-Ala-OH/HATU/DIPEA/ DCM; d) 2-nitrobenzenesulfonyl chloride/DIPEA/DCM; e) 1 eq. AcCI/DCM/MeOH; f) DEAD/Ph₃P/THF; g) PhSH/K₂CO₃/DMF; h) 3-nitrobenzylbromide/ Et₃N/DMF/THF; i) TBAF/THF

hydroxyl group of 1, however this would result in the 8-hydroxy group being present in all libraries prepared, and therefore limit the diversity achieved. To overcome this problem, we investigated the possibility of attaching a linker at an amino group on the 8-position of the final product, which would allow for functionalization at this position during the synthetic pathway.¹¹ The necessary support 6 was prepared from 4-hydroxybenzaldehyde and the acid stabile ArgoGelTM-OH resin¹² via modified Mitsunobu conditions,¹³ and the structure verified by gel-phase ¹³C NMR.¹⁴ The hydroxyprolinol derived scaffold was elaborated to an amine derivative by protecting 7^9 as the 2trimethylsilylethoxycarbonyl (Teoc)¹⁵ derivative (Scheme 2), followed by mesylation, azide displacement, and reduction¹⁶ to provide amine 8 in high yield. This material was then loaded onto support via a BH₃•pyridine¹⁷ mediated reductive amination, affording a quantitative yield of 9.¹⁸ The secondary amine of 9 was functionalized with an isocyanate to give urea 10, which could be cleanly cleaved from the support by treatment with TFA to provide 11 in quantitative yield and >90% purity.¹⁸

In order to follow the progress of subsequent reactions without detaching material from the support, isotopically enriched ¹⁵N-Alanine was used as the amino acid, which allowed convenient monitoring of the success of transformations by gel phase ¹⁵N NMR. This technique provided a convenient label for following the progress of the reaction sequence, as the amino-acid derived nitrogen is involved in the key ring forming reactions, and therefore exhibits significant differences in chemical shift. Furthermore, the sensitivity obtained was sufficient to gauge the



Scheme 2. a) Teoc-NHS; b) MsCl/CH₂Cl₂/pyridine; c) NaN₃/DMF/80 °C; d) SnCl₂/PhSH/Et₃N/MeCN e) 4 eq. p-tolyl isocyanate/DMF; f) CHCl₂CO₂H/CH₂Cl₂; g) CF₃CO₂H/5% Et₃SiH



Scheme 3. a) 1 M TBAF/THF; b) ¹⁵N-FMOC-Ala-OH/HATU/DIPEA/CH₂Cl₂; c) piperidine/DMF; d) 2-nitrobenzenesulfonyl chloride/DIPEA/CH₂Cl₂; e) CHCl₂CO₂H/CH₂Cl₂; f) 3 eq. Ph₃P-sulfonamide betaine (ref. 13)/5% DMF/ CH₂Cl₂; g) 0.5 M HSCH₂CO₂H/1 M DBU/DMF; h) 5 eq. thymine-1-acetic acid/HATU/DIPEA/ CH₂Cl₂; i) CF₃CO₂H/5% Et₃SiH success of failure of reactions, as by products were evident by the presence of multiple peaks when inappropriate

success or failure of reactions, as by-products were evident by the presence of multiple peaks when inappropriate reaction conditions were chosen.

After the Teoc protection of 10 (Scheme 3) was removed, the product was functionalized with ¹⁵N-FMOC-Ala-OH, and then deprotected to provide 12, which showed a single resonance at δ -350.6 ppm in the ¹⁵N NMR spectrum (referenced to formamide in DMSO-*d*₆). Introduction of the 2-nitrobenzene sulfonamide group, followed by removal of the DMT and gave 13 (¹⁵N NMR δ -275.5 ppm, doublet, *J*=79 Hz). Reaction under modified Mitsunobu conditions¹³ effected a clean cyclization to 14 (¹⁵N NMR δ -283.4 ppm, singlet), which was deprotected with mercaptoacetic acid/DBU to provide 15 (¹⁵N NMR δ -344.5 ppm). Functionalization of the unsubstituted nitrogen provided 16 (¹⁵N NMR δ -261.6 ppm), and cleavage of 15 and 16 gave the corresponding non-resin bound compounds 17 and 18. The yield of 18 (by mass) over the 12 steps from the commercially available support was 64%, with an estimated purity of 80% (by ¹H NMR), representing a true yield of ~50%, and an average chemical yield of 95%. This yield would most likely be unattainable with a traditional solution synthesis, and illustrates that reasonably long synthetic pathways are feasible for the synthesis of small molecule combinatorial libraries on solid support.

In conclusion, a synthetic pathway has been developed which provides functionalized piperazin-2-one ring systems in excellent yields. For demonstrative purposes, representative combinatorial library structures based upon a 3,4,8-trisubstituted 1,4-diazabicyclo[3.4.0]nonan-2-one scaffold have been prepared employing solid phase methodologies. This framework provides three variable positions for the introduction of diverse functionality, two via common electrophilic reagents, and one derived from commercially available FMOC-amino acids. The starting materials for the synthesis (*trans*-4-hydroxyproline and α -amino acids) determine the chirality of the final product, which eliminates stereochemical ambiguities. While a five-membered starting framework has been used to prepare a 6:5 fused ring system in this study, it should be noted that other functionalized heterocycles could presumably be prepared via this methodology. In particular, 6:6 fused systems would be accessible from azasugars such as the 1-deoxynojirimycin analogs.¹⁹ We have adapted this methodology to high throughput mode on an automated parallel array synthesizer, and are currently pursuing this strategy for the synthesis of large combinatorial libraries based upon this scaffold for screening in biological assays.

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REFERENCES AND NOTES

¹An, H.; Cummins, L. L.; Griffey, R. H.; Bharadwaj, R.; Haly, B. D.; Fraser, A. S.; Wilson-Lingardo, L.; Risen, L. M.; Wyatt, J. R.; Cook, P. D. J. Am. Chem. Soc. 1997 119, 3696-3708.

² Annual Reports in Combinatorial Chemistry and Molecular Diversity, Vol. 1; Moos, W. H.; Pavia, M. R.; Kay, B. K.; Ellington, A. D., Eds.; ESCOM: Leiden, 1997, and references cited therein.

³ Bunin, B. A.; Ellman, J. A. J. Am. Chem. Soc. 1992, 114, 10997-10998.

⁴ DeWitt, S. H.; Kiely, J. S.; Stankovic, C. J.; Schroeder, M. C.; Reynolds Cody, D. M.; Pavia, M. Proc. Natl. Acad. Sci. USA 1993, 90, 6909-6913.

⁵ Nefzi, A.; Ostresh, J. M.; Meyer, J.-P.; Houghten, R. A. Tetrahedron Lett. 1997, 38, 931-934.

⁶ a) Marx, M. A.; Grillot, A.-L.; Louer, C. T.; Beaver, K. A.; Bartlett, P. A. J. Am. Chem. Soc. **1997**, 119, 6153-6167. b) Bartlett, P. A.; Lewis, J. G.; Johnson, D. D.; Trump, R. P. Book of Abstracts, 213th ACS National Meeting, San Francisco, April 13-17 (1997), ORGN-273.

⁷Kozlowski, M. C.; Bartlett, P. A. J. Org. Chem. 1996, 61, 7681-7696.

⁸Fukuyama, T.; Jow, C.-K.; Cheung, M. Tetrahedron Lett. 1995, 36, 6373-6374.

⁹ Hebert, N.; Davis, P. W.; DeBaets, E. L.; Acevedo, O. L. Tetrahedron Lett. 1994, 35, 9509-9512.

¹⁰ Protogene Technologies, 4040 Fabian Way, Palo Alto, CA 94303.

¹¹ Swayze, E. E. Unpublished results. For related strategies, see: Boojamra, C. G.; Burow, K. M.; Thompson, L. A.; Ellman, J. A. J. Org. Chem. 1997, 62, 1240-1256 and Ngu, K.; Patel, D. V. Tetrahedron Lett. 1997, 38, 973-976.

¹² Argonaut Technologies, Inc. San Carlos, CA, (888) 598-1350.

¹³ Castro, J. L.; Matassa, V. G. J. Org. Chem. 1994, 59, 2289-2291.

¹⁴ Look, G. C.; Holmes, C. P.; Chinn, J. P.; Gallop, M. J. J. Org. Chem. 1994, 59, 7588-7570.

¹⁵ Shute, R. E.; Rich, D. H. Synthesis 1987, 346-349.

¹⁶ Bartra, M.; Romea, P.; Urpi, F.; Vilarrasa, J. Tetrahedron 1990, 46, 587-594.

¹⁷ a) Khan, N. M.; Arumugam, V.; Balasubramanian, S. *Tetrahedron Lett.* **1996**, *37*, 4819-4822. b) Bomann, M. D.; Guch, I. C.; DiMare, M. J. Org. Chem. **1995**, *60*, 5995-5996.

¹⁸ The yield of 9 was determined by spectrometric assessment of the amount of DMT cation released upon acid treatment of a sample of dried resin. The yield of 11 was determined by mass of cleaved product, and purity assessed by RP-HPLC employing an evaporative light scattering detector (SEDEX).

¹⁹ a) Baxter, Ellen W.; Reitz, A. B. J. Org. Chem. 1994, 59, 3175-3185. b) Enders, D.; Jegelka, U. Synlett. 1992, 999-1002. c) Furneaux, R. H.; Tyler, P. C.; Whitehouse, L. A. Tetrahedron Lett. 1993, 34, 3613-3616.

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